

Oxocyclostylidol, an Intramolecular Cyclized Oroidin Derivative from the Marine Sponge *Stylissa caribica*[§]

Achim Grube and Matthias Köck*

Alfred-Wegener-Institut für Polar- und Meeresforschung in der Helmholtz-Gemeinschaft, Am Handelshafen 12, D-27570 Bremerhaven, Germany

Received October 17, 2005

Pyrrole–imidazole alkaloids are widely distributed in marine sponges of the orders Halichondrida and Agelasida. Chemical investigation of the Caribbean sponge *Stylissa caribica* has led to the isolation of the first brominated pyrrole–imidazole alkaloid containing an oxidized pyrrole moiety. The isolation and structure elucidation of oxocyclostylidol (**1**) are discussed in detail.

Sponges of the order Halichondrida and Agelasida are well-known to be a rich source of pyrrole–imidazole alkaloids.¹ The structural diversity of this alkaloid family is related to oroidin (**2**),² which can react in different cyclization reactions.³ Products of dimerization reactions are the group of agelifेरins and scepтрins,⁴ while the group of axinellamines⁵ and massadine⁶ are oxidized dimerization products of oroidin (**2**). For example, in monomolecular cyclization reactions of oroidin (**2**), cyclooroidin (**3**)⁷ or stevensine⁸ is formed. In all of the described products the bromopyrrole moiety is not modified. In this paper we describe the first pyrrole–imidazole alkaloid from natural sources with a twice oxidized pyrrole moiety.

In our investigation of Caribbean organisms the sponge *Stylissa caribica* was examined. The sponge was extracted with a mixture of MeOH/CH₂Cl₂ (1:1), and the crude extract was partitioned by liquid/liquid extraction between *n*-hexane, *n*-BuOH, and H₂O. The resulting *n*-BuOH fraction was purified by Sephadex LH-20 chromatography. In an HPLC-HRMS screening of all fractions obtained from Sephadex LH-20 chromatography a compound with a mass pattern of *m/z* = 340/342 was detected, which was consistent with a bromine isotope pattern.

The molecular formula of oxocyclostylidol (**1**) was established by high-accuracy MS spectra. The structure of **1** was elucidated by UV, 2D NMR, and MS/MS spectra. Table 1 summarizes the 1D and 2D NMR data of **1**. The high-resolution mass of *m/z* 340.0032 indicated the molecular formula C₁₁H₁₁N₅O₃Br ([M + H]⁺) for **1**. Under MS/MS conditions the loss of 18 amu of the molecular ion suggested the presence of a hydroxyl group in **1**.

The carbon skeleton of the new compound **1** was elucidated using correlations from ¹H, ¹H-COSY, ¹H, ¹³C-HMBC, ¹H, ¹⁵N-HMBC, and 1,1-ADEQUATE experiments. The comparison of the ¹⁵N-chemical shifts of **1** and **2** indicated a change in the pyrrole ring but no changes in the imidazole moiety (see Figure 1). This is also confirmed by the identical number of correlations obtained in the ¹H, ¹⁵N-HMBC of **1** in comparison to oroidin (**2**). The ¹³C NMR signals at δ 110.9, 121.9, and 146.9 ppm are identical for the imidazole moiety in oroidin (**2**). Compared to **2** the signal for C-10 (**1**, 98.5; **2**, 126.4 ppm) is shifted to higher fields in **1**, whereas the signal for C-9 (**1**, 130.3; **2**, 112.7 ppm) is shifted to lower fields. These values supported a major change in the electron density of the double bond and indicated a directly bound heteroatom to C-9 or C-10. In combination with the correlation from H-10 to N-1 in

Table 1. NMR Data of Oxocyclostylidol (**1**) Recorded in DMSO-*d*₆^a

no.	δ _C ^b	δ _H	¹ H, ¹³ C-HMBC ^c	¹ H, ¹⁵ N-HMBC
1	(150)			
2	86.6			
2-OH		7.95 (s)	2, 3, 6	
3	144.5	7.75 (s)	<u>2</u> , <u>4</u> , 5, 9	1
4	119.5			
5	162.2			
6	165.4			
7	(104)	8.55 (d, <i>J</i> = 5.3 Hz)	2, 3, 6, 8, 9	
8	40.6	4.30 (dd, <i>J</i> = 2.2, 17.0 Hz)/ 4.09 (dd, <i>J</i> = 5.3, 17.0 Hz)	6, <u>9</u> , 10, 11	1, 7
9	130.3			
10	98.5	7.01 (s, br)	8, <u>9</u> , <u>11</u> , 12	1, 15
11	121.9			
12	110.9	6.85 (s, br)	8, 9, 10, <u>11</u> , 14	13, 15
13	(137)	12.31 (s, br)		
14	146.9			
15	(138)	12.44 (s, br)		
16	(59)	7.58 (s, br)		13, 15

^a ¹H and ¹³C chemical shifts [ppm] are referenced to the DMSO-*d*₆ signal (2.50 and 39.5 ppm, respectively). COSY correlations appear between H-7/H-8 (³*J*_{HH}) and H-8/H-10 (⁴*J*_{HH}). ^b For positions 1, 7, 13, and 15, δ_N is given in the δ_C column. ¹⁵N NMR spectra were not calibrated with an external standard. The δ value has an accuracy of about 1 ppm in reference to NH₃ (0 ppm). ^c HMBC correlations that also appear in the 1,1-ADEQUATE spectrum are underlined in the HMBC column. The correlations from H-3 to C-9, H-7 to C-3, H-8 to C-11, and H-12 to C-9 represent interactions over four bonds (⁴*J*_{CH}). The correlation from H-12 to C-8 represents an interaction over five bonds (⁵*J*_{CH}).

the ¹H, ¹⁵N-HMBC (see Supporting Information, Figure S1) this indicates the presence of an en-amine structure.

In contrast to other pyrrole–imidazole alkaloids the ¹H-signal of the pyrrole NH was missing and the ¹⁵N-chemical shift for the pyrrole nitrogen was shifted to higher fields (from 166 to 150 ppm) compared to oroidin (**2**). Three correlations to N-1 were observed in the ¹H, ¹⁵N-HMBC of **1** in contrast to oroidin (**2**), with only one correlation (see Supporting Information, Figure S1). The four remaining carbon atoms of the pyrrole moiety in **1** showed completely different chemical shifts (86.6, 119.5, 144.5, 162.2 ppm) from those obtained for monobrominated pyrrole alkaloids. The most downfield carbon atom at 162.2 ppm suggested the presence of an amide or ester function, whereas a ¹³C shift of 86.6 ppm is typical for a semi-aminale structure as observed in massadine (**4**).⁶ The remaining two carbon atoms at 119.5 and 144.5 ppm form the double bond in which the bromine atom is attached to the carbon atom at 119.5 ppm. The low-field shifted ¹H NMR signal of δ 144.5 and the ¹H, ¹⁵N-HMBC correlation between H-3 and N-1 (see Supporting Information, Figure S1) supported the existence of an α,β-unsaturated lactame. The structure of the pyrrole moiety was

[§] Presented at the 4th European Conference on Marine Natural Products, Paris, France, Sept 12–16, 2005 (Book of Abstracts, pp OC 09 and P 69).

* To whom correspondence should be addressed. Tel: +49-471-48311497. Fax: +49-471-48311425. E-mail: mkock@awi-bremerhaven.de.

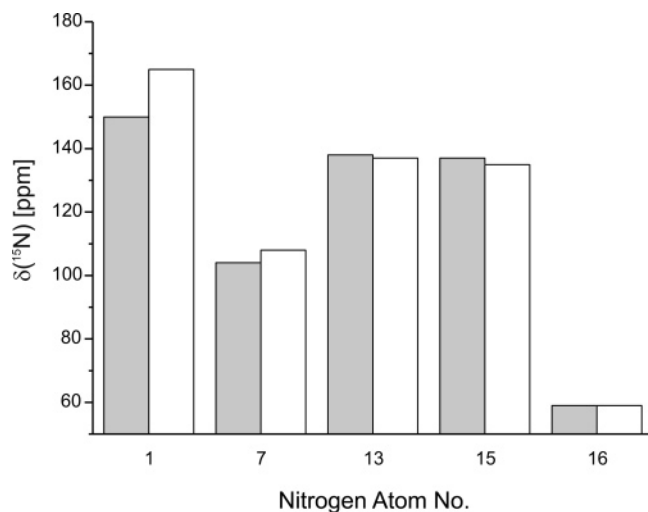


Figure 1. Graphical comparison of $\delta(^{15}\text{N})$ of **1** (gray bars) and **2** (white bars).

finally established by ^1H , ^{13}C -HMBC correlations between H-3 and C-2, C-4, and C-5 (see Supporting Information, Figure S2) and 1,1-ADEQUATE correlations from H-3 to C-2 and C-4 (see Supporting Information, Figure S3). The connection of the semi-aminal with the amide moiety is supported by a sharp ^1H NMR signal of the hydroxyl group indicating a hydrogen bond with the amide carbonyl. All these facts suggested a twice oxidized pyrrole system for oxocyclostylidol (**1**). This α -bromo, γ -hydroxy, α,β -unsaturated γ -butyrolactame system has not previously been observed in brominated pyrrole–imidazole alkaloids from natural sources. The configuration of the double bond between C-9/C-10 was assigned by ROESY spectra with different mixing times (150 and 200 ms). A strong ROE between H-8/H-12 and a missing ROE between H-10/H-12 indicated an *E* configuration of the double bond.

While semi-aminal structures are in general unstable, this structural element is known to be stable in neutral media (no hydrolysis products were detected in HPLC) in pyrrole–imidazole alkaloids. Massadine (**4**) and the axinellamines⁵ are stable compounds containing a semi-aminal structure. The structure of **1** included a vinylogous system between N-1 and the guanidine group, which is in accordance with the long-wave absorption maximum in the UV spectrum (320 nm). The absorption maximum at 263 nm is in the same region as for known pyrrole–imidazole alkaloids.

To verify the proposed structure of **1**, COCON calculations⁹ were carried out. For the experimental data set COCON generated four

Chart 1. Structural Formulas of Oxocyclostylidol (**1**), Oroidin (**2**), Cyclooroidin (**3**), and Massadine (**4**)

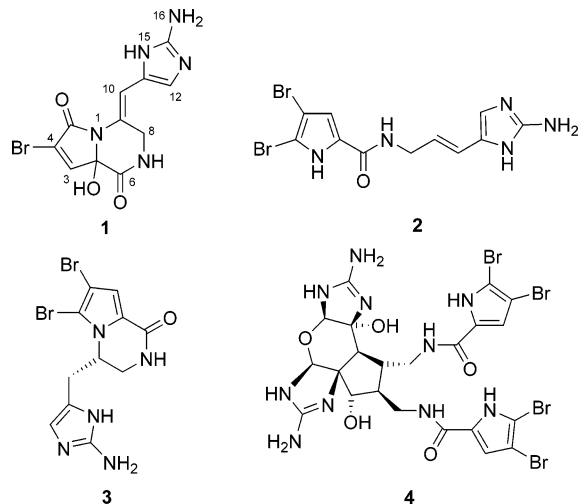
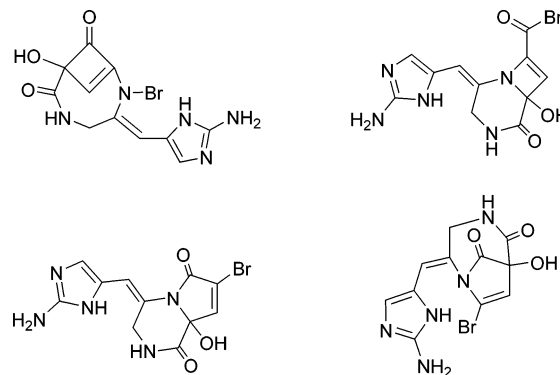


Chart 2. Four Structural Proposals for Oxocyclostylidol (**1**) Generated by COCON^a



^a All experimental data including correlations from ^1H , ^{15}N -HMBC and 1,1-ADEQUATE were used as input for this COCON calculation.

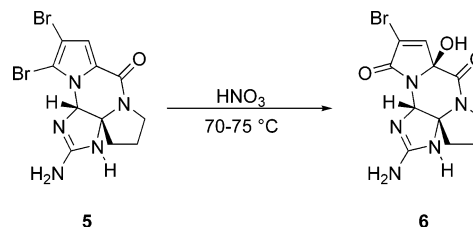


Figure 2. Reaction of dibromophakellin (**5**) with dilute nitric acid to the oxidized product **6**.¹¹

structural proposals (see Chart 2), which included the structure of oxocyclostylidol (**1**). The other three proposals can be neglected because two of them represent strained structures and the last is a carboxylic acid bromide. When running the COCON calculations without the 1,1-ADEQUATE data and the ^1H , ^{15}N -HMBC data, the number of possible structures increases rapidly.⁹ The calculation without the ^1H , ^{15}N -HMBC data led to more than 50 000 structural proposals. This number increases to more than 150 000 structural proposals if the 1,1-ADEQUATE data were also not used. These numbers demonstrated the importance of the ^1H , ^{15}N -HMBC data for the unambiguous structure elucidation of oxocyclostylidol (**1**).¹⁰

To the best of our knowledge, oxocyclostylidol (**1**) is the first pyrrole–imidazole alkaloid from natural sources that includes an oxidized pyrrole moiety. The carbon–nitrogen skeleton of **1** is identical to cyclooroidin (**3**).⁷ Biosynthetically, oxocyclostylidol (**1**) could be derived from an intramolecular cyclized oroidin (**2**) that is twice oxidized under loss of bromine in position 5. This is supported by synthetic approaches to oxidized bromopyrrole moieties. This functionality is known from the reaction of dibromophakellin (**5**) with dilute nitric acid¹¹ and as a byproduct in the biomimetic synthesis of dibromophakellin (**5**)¹² (see Figure 2).

The new structure of oxocyclostylidol (**1**) includes some interesting functional groups that could induce biological or pharmacological effects, which are still under investigation. Oxocyclostylidol (**1**) was tested against several pathogenic bacteria, fungi, and cultures of mice fibroblasts, but showed only minor activity in these tests. Determination of the absolute configuration will be carried out by synthesis.

Experimental Section

General Experimental Procedures. Optical rotation was measured with a Perkin-Elmer 241 MC polarimeter at 23 °C. CD spectra were recorded with a Jasco J-810 system. UV spectra were recorded with a DAD (Agilent) during HPLC analysis. ^1H NMR and ^{13}C NMR spectra were recorded on Bruker Avance 400 and 600 NMR spectrometers. All experiments were measured at 298 or 300 K (sample concentration: 9.5 mg/600 μL). The DQF- ^1H , ^1H -COSY, ^1H , ^{13}C -HSQC, ^1H , ^{13}C -HMBC, ^1H , ^{15}N -HSQC, ^1H , ^{15}N -HMBC, 1,1-ADEQUATE, and ^1H , ^1H -ROESY experiments were carried out using standard parameters.

HPLC-MS analyses were performed with an Agilent 1100 HPLC system and a Bruker Daltonics microTOF_{LC} mass spectrometer. Separation was achieved by a Waters XTerra RP₁₈ column (3.0 × 150 mm, 3.5 μm) applying a MeCN/0.01% HCOOH (in water) gradient (0 min: 10% MeCN/90% HCOOH (0.01%); 30 min: 60% MeCN/40% HCOOH (0.01%)) with a flow rate of 0.4 mL/min. ESIMS/MS spectra were recorded with an Esquire 3000+ ion trap (Bruker Daltonics).

Animal Material. The sponge *Stylissa caribica* was collected by scuba at Little San Salvador in the Bahamas (74 ft depth, July 2000). The samples were immediately frozen after collection and kept at -20 °C until extraction. The sponge material was identified as previously described.¹³

Extraction and Isolation. The freeze-dried sponge samples of *S. caribica* (94.7 g) were crushed with a mill and extracted at RT exhaustively in a 1:1 mixture of CH₂Cl₂/MeOH. The orange-colored crude extract of *S. caribica* was partitioned between *n*-hexane (4 × 400 mL) and MeOH (300 mL). The MeOH extract was then partitioned between *n*-BuOH (3 × 500 mL) and H₂O (300 mL). The resulting *n*-BuOH (15.9 mg) phase from the solvent-partitioning scheme was purified by gel chromatography on Sephadex LH-20 (Pharmacia) using MeOH as mobile phase. Final purification of the isolated compound was achieved by preparative RP₁₈ HPLC on a Kromasil RP₁₈ column (16 × 250 mm, 10 μm) applying a MeCN/TFA (0.1% in water) gradient to afford **1** (29.5 mg, 0.03% of dry weight).

Oxocystylidol (1): yellow powder; [α]_D²³ -12 (c 1, MeOH); UV (DAD) λ_{max} 224, 263, and 320 nm; CD (MeCN) λ (Δε) 208 (-6.1), 235 (1.7), 270 (1.5), 304 (-0.4) nm; HPLC/HR(+)-ESIMS *t*_R = 3.1 min, *m/z* 340.0032 [M + H]⁺ (calcd for C₁₁H₁₁N₅O₃⁷⁹Br 340.0040), Δ*m* = 1.8).

Acknowledgment. The sponge collection was carried out by M. Assmann during a scientific expedition to the Bahamas in 2000. During this time the project was sponsored by the DFG (Ko1314/3-1 to 3-4). We acknowledge the support of J. R. Pawlik (University of North Carolina at Wilmington), who gave members of the Köck research group the opportunity to participate in the scientific sojourns to the Bahamas in the years 1998, 1999, 2000, 2001, and 2003. We further

thank E. Lichte for performing preparative HPLC analysis, F. Sasse (GBF, Braunschweig) for biological testing, and T. Lindel (LMU München) for measuring the CD spectra.

Supporting Information Available: NMR spectra (¹H, ¹³C-HMBC, ¹H, ¹⁵N-HMBC, and 1,1-ADEQUATE) and CD spectrum in acetonitrile of oxocystylidol (**1**). This material is available free of charge via the Internet at <http://pubs.acs.org>.

References and Notes

- (1) Faulkner, D. J. *Nat. Prod. Rep.* **2002**, *19*, 1–48.
- (2) (a) Forenza, S.; Minale, L.; Riccio, R.; Fattorusso, E. *J. Chem. Soc. D* **1971**, 1129–1130. (b) Garcia, E. E.; Benjamin, L. E.; Fryer, R. I. *J. Chem. Soc., Chem. Commun.* **1973**, 78–79. (c) Walker, R. P.; Faulkner, D. J.; van Engen, D.; Clardy, J. *J. Am. Chem. Soc.* **1981**, *103*, 6772–6773.
- (3) Al Mourabit, A.; Potier, P. *Eur. J. Org. Chem.* **2001**, 237–243.
- (4) Keifer, P. A.; Schwartz, R. E.; Koker, M. E. S.; Hughes, R. G., Jr.; Rittschof, D.; Rinehart, K. L. *J. Org. Chem.* **1991**, *56*, 2965–2975; errata 5736, 6728.
- (5) Urban, S.; De Almeida Leone, P.; Carroll, A. R.; Fechner, G. A.; Smith, J.; Quinn, R. J.; Hooper, J. N. A. *J. Org. Chem.* **1999**, *64*, 731–735.
- (6) Nishimura, S.; Matsunaga, S.; Fusetani, N.; Shibasaki, M.; Suzuki, K.; Furihata, K.; van Soest, R. W. M. *Org. Lett.* **2003**, *5*, 2255–2257.
- (7) Fattorusso, E.; Tagliatalata-Scafati, O. *Tetrahedron Lett.* **2000**, *41*, 9917–9922.
- (8) Albizzati, K. F.; Faulkner, D. J. *J. Org. Chem.* **1985**, *50*, 4163–4164.
- (9) (a) Lindel, T.; Junker, J.; Köck, M. *J. Mol. Model.* **1997**, *3*, 364–368. (b) Lindel, T.; Junker, J.; Köck, M. *Eur. J. Org. Chem.* **1999**, 573–577. (c) Köck, M.; Junker, J.; Maier, W.; Will, M.; Lindel, T. *Eur. J. Org. Chem.* **1999**, 579–586.
- (10) Köck, M.; Junker, J.; Lindel, T. *Org. Lett.* **1999**, *1*, 2041–2044.
- (11) Sharma, G.; Magdoff Fairchild, B. *J. Org. Chem.* **1977**, *42*, 4118–4124.
- (12) Foley, L. H.; Büchi, G. *J. Am. Chem. Soc.* **1982**, *104*, 1776–1777.
- (13) Grube, A.; Lichte, E.; Köck, M. *J. Nat. Prod.* **2006**, *69*, 125–127.

NP050408F